

**Supplementary information for the article:**

Ljubic, V.; Milosevic, M.; Cvetkovic, S.; Stojanovic, M.; Novovic, K.; Dinic, M.; Popovic, M. The New Exopolysaccharide Produced by the Probiotic Strain *L. Reuteri* B2: Extraction, Biological Properties, and Possible Application for Ni<sup>2+</sup> Ion Removal from the Contaminated Water. *Biomass Conversion and Biorefinery* 2022. <https://doi.org/10.1007/s13399-022-03292-5>.



## SUPPLEMENTARY MATERIAL

### **Partial characterization and biological properties of the novel exopolysaccharide produced by a probiotic strain *L. reuteri* B2**

Verica Ljubic<sup>1</sup>, Milena Milosevic<sup>1</sup>, Slobodan Cvetkovic<sup>1</sup>, Marijana Stojanovic<sup>2</sup>, Katarina Novovic<sup>3</sup>, Miroslav Dinic<sup>3</sup>, and Mina Popovic<sup>1\*</sup>

<sup>1</sup> University of Belgrade, Institute of Chemistry, Technology, and Metallurgy, National Institute of Republic of Serbia, Njegoseva 12, 11000 Belgrade, Serbia, corresponding author

<sup>2</sup> Institute of Virology, Vaccines, and Sera, Torlak, Vojvode Stepe 458, 11000 Belgrade, Serbia

<sup>3</sup> Institute of Molecular Genetics and Genetic Engineering, University of Belgrade, Vojvode Stepe 444a, 11042 Belgrade, Serbia.

## **2.1 Bacteria and culture conditions**

### **2.1.1 Strain identification.**

Using the universal primers (UNI16Sfw and UNI16Srev) [1], the representative isolates were identified by 16S rDNA sequencing. Amplification was carried out in a thermal cycler (Applied Biosystems, ThermoFisher Scientific) and DNA fragments were amplified as follows: initial denaturation at 94 °C for 5 min, followed by 30 cycles consisting of denaturation at 94 °C for 1 min, annealing at 55°C for 1 min, and polymerization at 72°C for 1 min, and a final extension at 72°C for 7 min. The expected length was 1549 bp. Aliquots (5 µl) of the amplified products were subjected to electrophoresis in 1% agarose gel (ThermoFisher Scientific) in TAE buffer (40 mM Tris acetate, 1 mM EDTA, pH 8.2). Gels were stained with ethidium bromide (500 ng/mL) and visualized under UV light (BioDoc Analyze). All amplicons were eluted and purified using GeneJet PCR Purification Kit (ThermoScientific) by following the manufacturer's protocol. The PCR products that we obtained were sequenced by the MacroGen Sequencing Service (MacroGen, Amsterdam, The Netherlands) and analyzed by using BLAST algorithm (<http://www.ncbi.nlm.nih.gov/index.html>). Selected isolates were identified as follows: isolate B2 - *Lactobacillus reuteri*, isolate H10 - *Lactobacillus murinus*, and isolate J7 - *Klebsiella oxytoca* [2]. The most numerous colonies belong to isolate B2, hence it was chosen for further characterization as a potential source for exopolysaccharide (EPS) production.

### **3.1.2 Characterization of EPS isolated from *L. reuteri* B2**

## <sup>1</sup>H NMR spectra of new EPS from *L. reuteri* B2

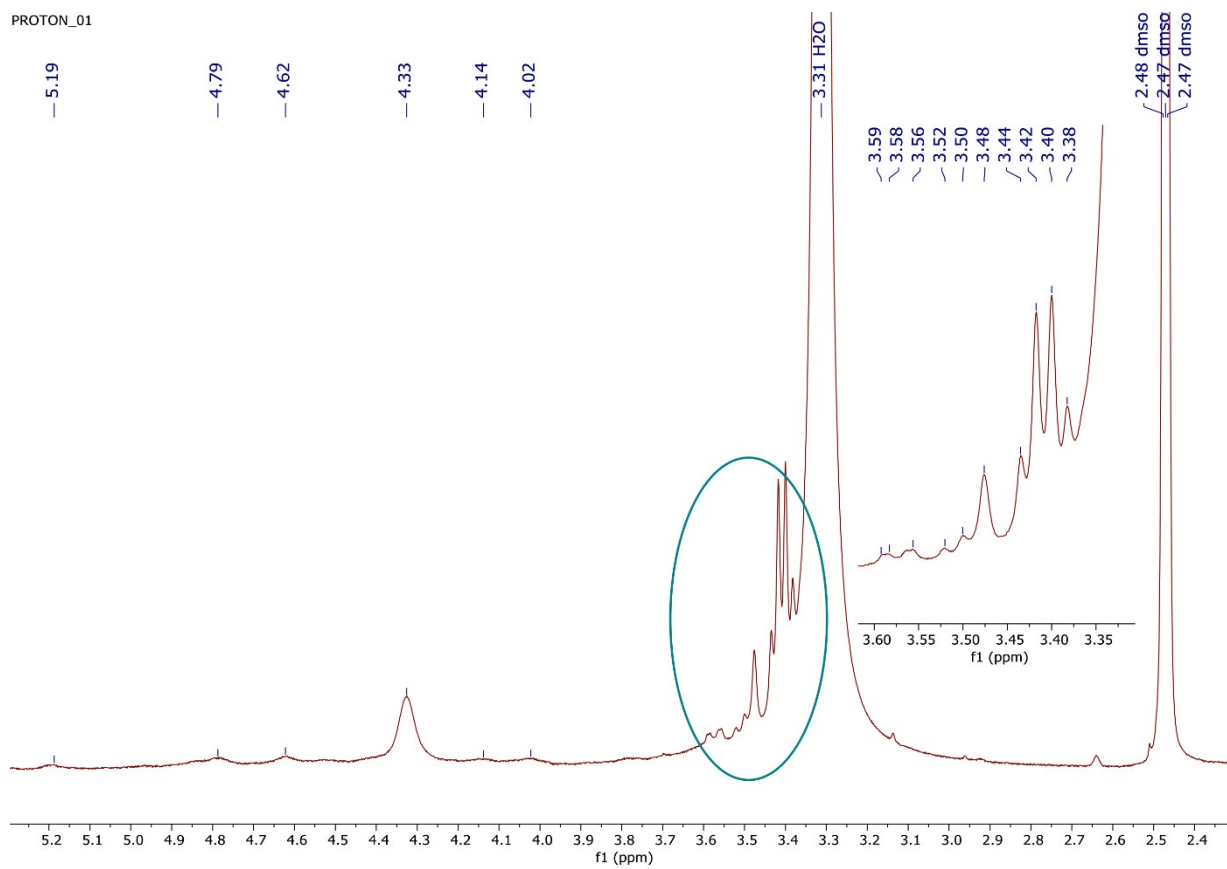


Fig S1. <sup>1</sup>H NMR spectra of EPS from *L. reuteri* B2 recorded on a Bruker Avance III 500 spectrometer in DMSO-d<sub>6</sub>

### 3.2.4 Determination of point zero charges (PZC) of EPS from *L. reuteri* B2

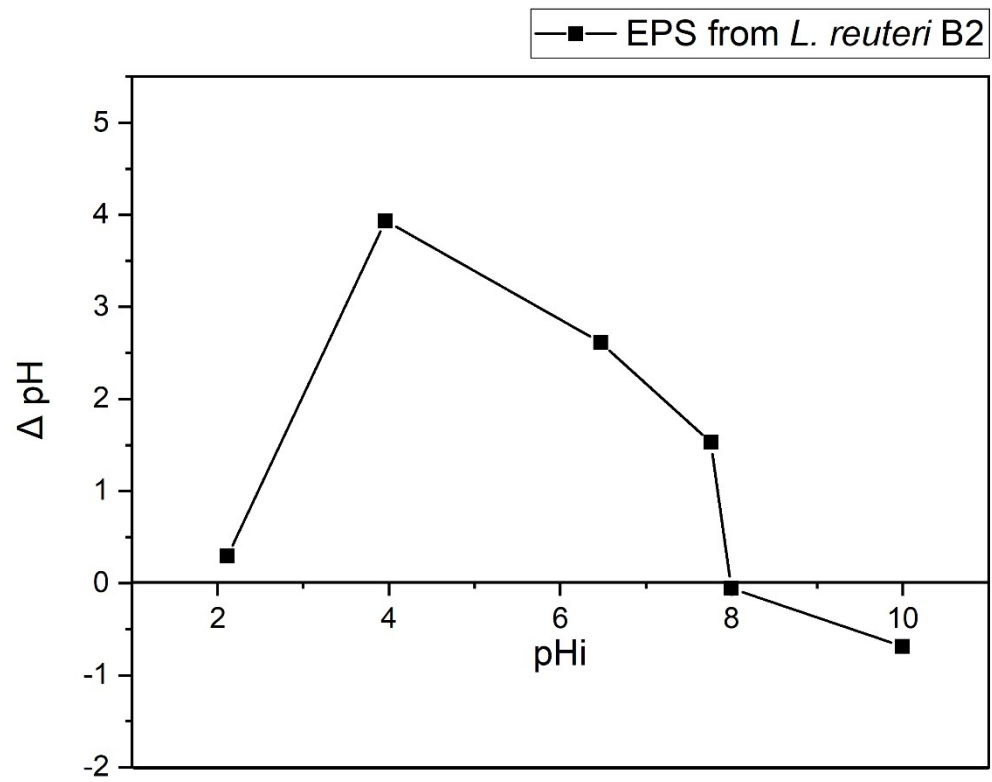
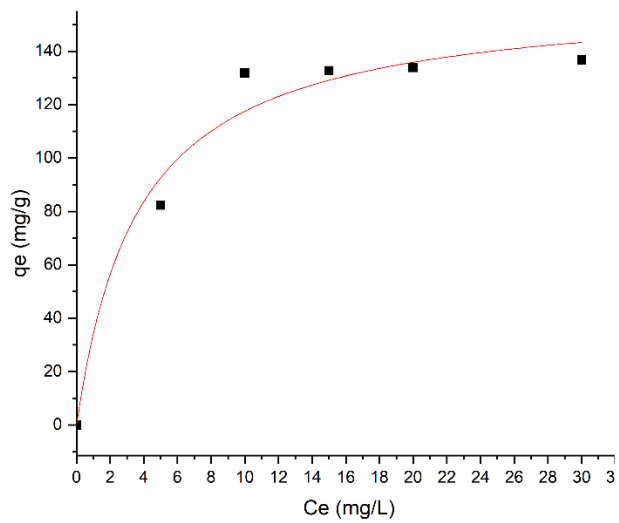


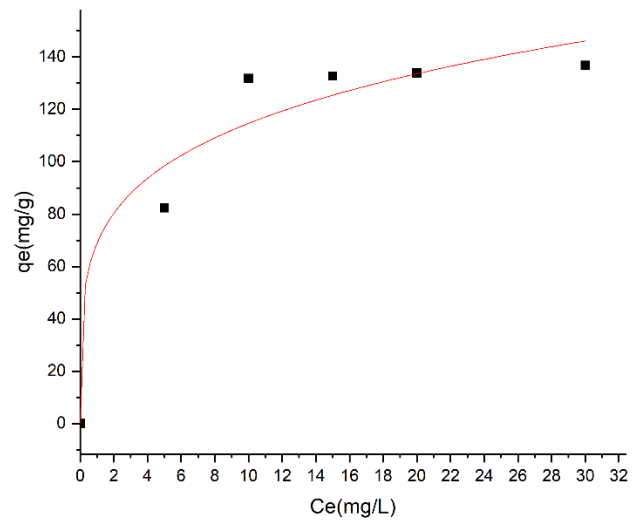
Fig. S2. Point of zero charge of EPS from *L. reuteri* B2

### 3.4.1 Isotherm study

a)

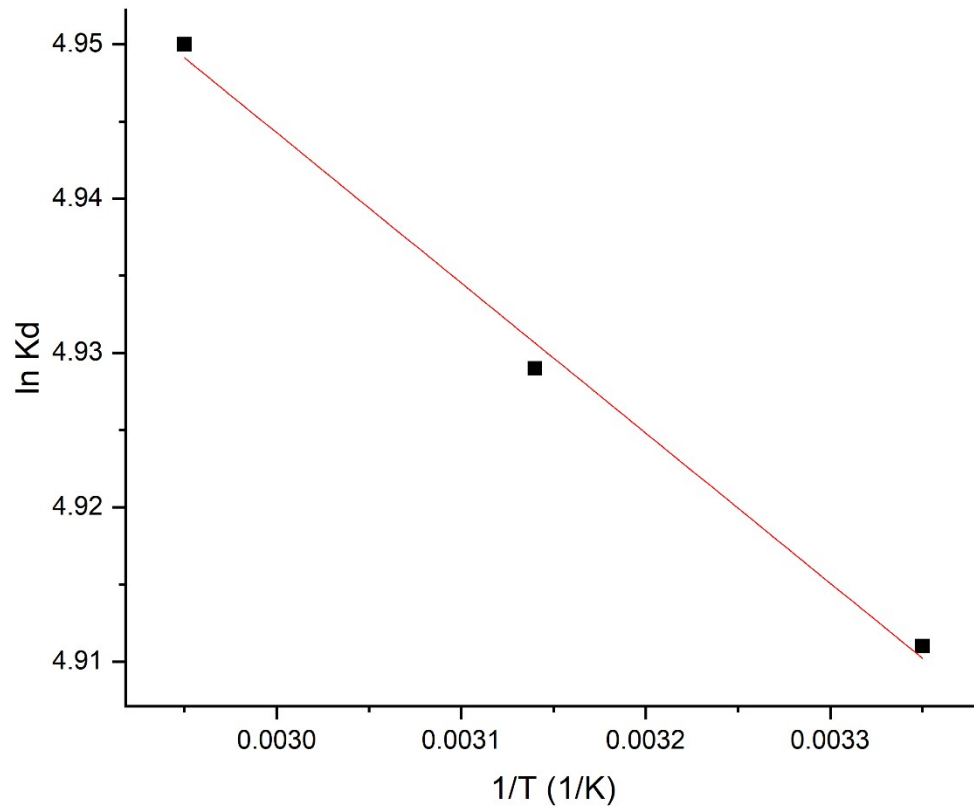


b)



**Fig. S3.** Isotherm study, Langmuir a), and Freundlich b) isotherms for adsorption of  $\text{Ni}^{2+}$  ions on EPS isolated from *L. reuteri* B2, equilibrium experiments were performed for different  $\text{Ni}^{2+}$  concentrations (0-35 mg/L) at 25 °C, for 60 min at pH 8.0.

### 3.4.2 Thermodynamic study



**Fig. S4.** Thermodynamic study of the adsorption for  $\text{Ni}^{2+}$  ions on EPS isolated from *L. reuteri* B2, equilibrium experiments were performed for  $\text{Ni}^{2+}$  concentrations 10 mg/L at different temperatures (298,318 and 338 K) for 60 min at pH 8.0.

## References

1. Jovic B, Begovic J, Lozo J, et al (2009) Dynamics of sodium dodecyl sulfate utilization and antibiotic susceptibility of strain *Pseudomonas* sp. ATCC19151. *Arch Biol Sci* 61:159–164. <https://doi.org/10.2298/abs0902159j>
2. Popović M, Stojanović M, Veličković Z, et al (2021) Characterization of potential probiotic strain, *L. reuteri* B2, and its microencapsulation using alginate-based biopolymers. *Int J Biol Macromol* 183:423–434. <https://doi.org/10.1016/j.ijbiomac.2021.04.177>